This article was downloaded by: On: 22 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Asian Natural Products Research

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713454007

Two new compounds from the roots of *Ligusticum chuanxiong*

Xin-Liang Chang^a; Zhi-Yong Jiang^a; Yun-Bao Ma^a; Xue-Mei Zhang^a; Karl W. K. Tsim^b; Ji-Jun Chen^a ^a tate Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming, Yunnan, China ^b Department of Biology and Center for Chinese Medicine, Hong Kong University of Science and Technology, Clear Water Bay Road, Hong Kong, China

To cite this Article Chang, Xin-Liang , Jiang, Zhi-Yong , Ma, Yun-Bao , Zhang, Xue-Mei , Tsim, Karl W. K. and Chen, Ji-Jun(2009) 'Two new compounds from the roots of *Ligusticum chuanxiong*', Journal of Asian Natural Products Research, 11: 9, 805 – 810

To link to this Article: DOI: 10.1080/10286020903071068 URL: http://dx.doi.org/10.1080/10286020903071068

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



Two new compounds from the roots of Ligusticum chuanxiong

Xin-Liang Chang^a, Zhi-Yong Jiang^a*, Yun-Bao Ma^a, Xue-Mei Zhang^a, Karl W.K. Tsim^b and Ji-Jun Chen^a*

^aState Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, Yunnan, China; ^bDepartment of Biology and Center for Chinese Medicine, Hong Kong University of Science and Technology, Clear Water Bay Road, Hong Kong, China

(Received 15 April 2009; final version received 26 May 2009)

Two new compounds, named ligusticoside A (1), a novel phthalide derivative with a lactone ring, and 4-pentylcyclohex-3-ene- 1α ,2 β -diol (2), were isolated from the rhizomes of *Ligusticum chuanxiong*. Their structures were determined by spectral methods (MS, IR, UV, 1D and 2D NMR). An X-ray diffraction experiment was performed to confirm the structure of compound 1.

Keywords: *Ligusticum chuanxiong*; phthalide derivatives; ligusticoside A; 4-pentylcyclohex-3-ene- 1α , 2β -diol

1. Introduction

Ligusticum chuanxiong Hort., a member of the Umbelliferae family, is mainly cultivated in Sichuan Province of China, and its root is an important traditional Chinese medicine being used to treat headaches, anemia, and irregular menstrual cycles [1]. Previous chemical analyses on the roots of L. chuanxiong have reported the occurrence of phthalides, dimeric phthalides, terpenes, alkaloids, and organic acids [2-10]. In order to further understand its chemical diversities, the water extract of L. chuanxiong was phytochemically investigated to afford two new compounds, ligusticoside A (1) and 4-pentylcyclohex-3-ene-1 α ,2 β -diol (2) (Figure 1). This paper describes the isolation and structural elucidation of the two new compounds. The structure of ligusticoside A(1) was also confirmed by an X-ray diffraction experiment.

2. Results and discussion

Compound 1, $[\alpha]_{\rm D}^4$ +72.33 (*c* = 0.26, MeOH), was obtained as a colorless prism (MeOH). The molecular formula was deduced to be C₁₈H₂₈O₈ by negative HR-ESI-MS at m/z 371.1704 [M - 1]⁻, indicating the presence of five degrees of unsaturation in the molecule. The IR spectrum displayed absorptions ascribable to hydroxyl (3486 cm^{-1}) and ester carbonyl (1752 cm^{-1}) functionalities. The ¹H NMR spectrum (Table 1) exhibited two olefinic proton signals at $\delta_{\rm H}$ 5.88 (dd, J = 8.6, 7.8 Hz), 5.69 (br d, $J = 8.6 \,\mathrm{Hz}$), a triplet methyl group at $\delta_{\rm H}$ 0.82 (t, $J = 6.5 \,\rm{Hz}$), as well as an anomeric proton at $\delta_{\rm H}$ 5.06 (1H, d, $J = 7.8 \,\mathrm{Hz}$), indicating that there was a β -configuration sugar moiety in the molecule. The ¹³C NMR (DEPT) spectrum (Table 1) showed 18 carbon signals: one ester carbonyl group ($\delta_{\rm C}$ 177.5), one

ISSN 1028-6020 print/ISSN 1477-2213 online © 2009 Taylor & Francis DOI: 10.1080/10286020903071068 http://www.informaworld.com

^{*}Corresponding authors. Email: chenjj@mail.kib.ac.cn; jiangzy@mail.kib.ac.cn



Figure 1. The structures of compounds 1 and 2.

quaternary carbon ($\delta_{\rm C}$ 85.7), four methines including two olefinic carbons at $\delta_{\rm C}$ 128.8 and 122.4, and one oxygen-bearing methine at $\delta_{\rm C}$ 81.2, and five aliphatic methylenes at $\delta_{\rm C}$ 32.5, 31.6, 28.0, 22.8, and 22.6, along with one glucopyranosyl unit at $\delta_{\rm C}$ 99.1, 74.8, 78.8, 72.2, 78.3, and 63.8 which was identified by the comparison of its NMR spectral data with those of a β -D-glucopyranosyl moiety of (4*S*)-*p*-menth-1-ene-4,7-diol 4-*O*- β -D-glucopyranoside [11].

| | 1 | | 2 | |
|-----|---|----------------------|---|--------------------------|
| No. | ¹ H ($\delta_{\rm H}$, mult., J in Hz) | $^{13}C(\delta_{C})$ | ¹ H ($\delta_{\rm H}$, mult., J in Hz) | $^{13}C(\delta_{\rm C})$ |
| 1 | _ | 177.5 (s) | 3.54-3.49 (m) | 73.8 (d) |
| 2 | _ | - | 3.92-3.90 (m) | 73.6 (d) |
| 3 | 4.83 (br s) | 81.2 (d) | 5.27 (br s) | 123.7 (d) |
| 4 | | 85.7 (s) | _ | 141.8 (s) |
| 5 | 3.65 (d, 7.3) | 48.8 (d) | 2.07-2.04 (m) | 28.1 (t) |
| 6 | 5.88 (dd, 8.6, 7.3) | 124.4 (d) | 1.90–1.84 (m), | 29.8 (t) |
| | | | 1.64 - 1.57 (m) | |
| 7 | 5.69 (br d, 8.6) | 128.8 (d) | 1.98–1.95 (t, 7.5) | 37.9 (t) |
| 8 | 2.53-2.40 (m) | 31.6 (t) | 1.46-1.35 (m) | 28.4 (t) |
| 9 | 1.98–1.81 (m) | 28.0 (t) | 1.35–1.24 (overlapped) | 32.7 (t) |
| 10 | 2.07-2.04 (m), | 22.6 (t) | 1.35 - 1.24 (overlapped) | 23.6 (t) |
| | 1.44 - 1.40 (m) | | | |
| 11 | 1.24 (m) | 32.5 (t) | 0.89 (t, 6.9) | 14.4 (q) |
| 12 | 1.24 (m) | 22.8 (t) | | |
| 13 | 0.82 (t, 6.5) | 14.3 (q) | | |
| 1' | 5.06 (d, 7.8) | 99.1 (d) | | |
| 2' | 4.04-3.92 (overlapped) | 74.8 (d) | | |
| 3' | 4.21 (t, 8.7) | 78.8 (d) | | |
| 4′ | 4.04-3.92 (overlapped) | 72.2 (d) | | |
| 5' | 4.04–3.92 (overlapped) | 78.3 (d) | | |
| 6′ | 4.53 (br d, 10.4), 4.15–4.10 (m) | 63.8 (t) | | |

Table 1. ¹H and ¹³C NMR spectral data of compounds 1 and 2 in $C_5D_5N^a$.

^aAssignments were performed on the HSQC and HMBC spectra.

Except for three degrees of unsaturation due to one carbonyl group, one olefin, and one sugar, the remaining two degrees of unsaturation could be assigned to a bicyclic system in compound 1, which was substantiated by 2D NMR (¹H–¹H COSY, HSQC, HMBC) analyses. The correlated proton signals H-5/H-6, H-6/H-7, H-7/H-8, and H-8/H-3 in the ¹H-¹H COSY spectrum of compound 1 (Figure 2), together with the correlations H-3/C-4, H-5/C-4, H-8/C-3, H-8/C-7, and H-5/C-6 in the HMBC spectrum (Figure 2), suggested the existence of a ring between C5-C6-C7-C8-C3-C4. The presence of n-pentyl (C-9 to C-13) could also be deduced by the ¹H–¹H COSY and HMBC analyses (Figure 2). The correlations between H-3 ($\delta_{\rm H}$ 4.83), H-5 ($\delta_{\rm H}$ 3.65) and C-1 ($\delta_{\rm C}$ 177.5), H-9 ($\delta_{\rm H}$ 1.98–1.81), H-1' ($\delta_{\rm H}$ 5.06) and C-4 ($\delta_{\rm C}$ 85.7) in the HMBC spectrum (Figure 2) established that C-3 and C-5 were connected through an ester group and the β -D-glucopyranosyl and *n*-pentyl units were attached at C-4. Thus, compound 1 was characterized as shown in Figure 1.

The compound **1** is a new rearranged phthalide derivative featuring quaternary carbon glycoside. To further confirm the structure of compound **1**, an X-ray diffraction experiment was performed (Figure 3) through which the structure of compound **1** was corroborated as depicted in Figure 1.

Compound 2 was obtained as a colorless oil, $[\alpha]_{D}^{23} - 2.01$ (c = 0.74, MeOH). The molecular formula was determined to be $C_{11}H_{20}O_2$ by positive HR-ESI-MS at m/z 207.1353 [M+Na]⁺, with two degrees of unsaturation. The IR spectrum showed the presence of the hydroxyl group at $3420 \,\mathrm{cm}^{-1}$. In the ¹H NMR spectrum, one triplet methyl signal at $\delta_{\rm H}$ 0.89 (3H, t, J = 6.9 Hz, H-11) was observed, as well as an olefinic proton signal at $\delta_{\rm H}$ 5.27 (1H, br s). The ¹³C NMR (DEPT) spectrum (Table 1) suggested that there were two olefinic carbons, two oxygenated methines, six aliphatic methylenes, and one methyl in compound 2. In consideration of two degrees of unsaturation in compound 2, it could be deduced that compound 2 contained one ring, which was supported by the HMBC correlations (Figure 2) of H-2/C-1, H-2/C-3, H-5/C-4, H-6/C-1, and H-6/C-5, suggesting the existence of a C1-C2-C3-C4-C5-C6 (cyclohex-3ene-1,2-diol) ring. The HMBC spectrum (Figure 2), combined with the ${}^{1}H - {}^{1}H$



Figure 2. ${}^{1}H^{-1}H COSY$ (—) and selected HMBC (H \rightarrow C) correlations of compounds 1 and 2.



Figure 3. The X-ray crystal structure of compound 1.

COSY experiment (Figure 2), also established the presence of an *n*-pentyl group in the molecule of compound 2. The assignment of the *n*-pentyl group attached at C-4 was performed through the HMBC correlations between H-7, 8 and C-4. Thus, compound 2 was determined as 4pentylcyclohex-3-ene-1,2-diol. In order to interpret the relative stereochemistry of compound 2, a ROESY experiment was correlation conducted. The absent between H-1, assumed to be β -oriented, and H-2 in the ROESY spectrum demonstrated that H-1 and H-2 were in antarafacial. Thus, the structure of compound 2 was deduced as 4-pentylcyclohex-3-ene-1 α ,2 β -diol (2).

3. Experimental

3.1 General experimental procedures

Optical rotations were carried out on a HORIBA SEPA-300 high-sensitive polarimeter. IR spectra were recorded on a Bio-Rad FTS-135 spectrometer with KBr pellets, ν in cm⁻¹. 1D and 2D NMR experiments were performed on a Bruker-

AM-400 (¹H and ¹³C at 400 and 100 MHz, respectively) spectrometer, with TMS as the internal reference, J in Hz. Mass spectra were recorded on a VG-Auto-Spec-3000 instrument. Silica gel (200-300 mesh) for column chromatography was obtained from Qingdao Marine Chemical Factory (Qingdao, China); D₁₀₁ macroporous resins were obtained from Tianjin Pesticide Chemical Company (Tianjin, China); MCI gel CHP-20P (70-150 µm) was purchased from Mitsubishi Chemical Corporation (Tokyo, Japan); Lichrospher Rp-18 gel was purchased from Merck Chemical Ltd (40-63 µm; Darmstadt, Germany). Detection was performed by silica gel TLC on which 10% H₂SO₄ in EtOH was sprayed, followed by heating.

3.2 Plant material

The roots of *L. chuanxiong* used in this experiment were collected in 9 October 2004 in Sichuan Province, China, and identified as *L. chuanxiong* Hort. by Prof. Dr Li-Gong Lei from the Kunming Institute of Botany, Chinese Academy of

Sciences. A voucher specimen (No. 2004-10-09) has been deposited in the Kunming Institute of Botany, Chinese Academy of Sciences.

3.3 Extraction and isolation

The air-dried and powdered roots of L. chuanxiong (53 kg) were extracted with boiling water. The extract was obtained after evaporation of the solvent under vacuum, which was suspended in water and partitioned between EtOAc and *n*-BuOH, respectively, to afford the EtOAc and n-BuOH fractions. The n-BuOH fraction (1.6 kg) was applied onto a D_{101} macroporous resin column and eluted with H₂O, 20% EtOH, 50% EtOH, and 95% EtOH to give the corresponding fractions. The fraction eluted by 50% EtOH (400 g) was separated on a silica gel column $(CHCl_3 - MeOH - H_2O, 70:30:5 v/v)$ to give six fractions (1-6). Fraction 5 (16 g)was performed on an MCI CHP-20P gel column (MeOH-H₂O, 80:20) and subsequently purified on an Rp-18 column with an eluent of MeOH-H₂O (80:20) to yield compound 1 (100 mg). Fraction 3 (30 g) was subjected to a silica gel column (400 g, 200-300 mesh) eluted with CHCl₃-MeOH-H₂O (85:15:2 v/v) to furnish three subfractions (3.1-3.3). Subfraction 3.1 (1.5 g) was purified on an Rp-18 column (MeOH $-H_2O$, 85:15) followed by Sephadex LH-20 (MeOH) to obtain compound 2 (50 mg).

3.3.1 Ligusticoside A (1)

A colorless prism (MeOH); $[\alpha]_D^{14} + 72.33$ (c = 0.26, MeOH); UV λ_{max} (MeOH) nm (log ε): 218 (3.26), 197 (3.06), 193 (3.06); IR (KBr) ν_{max} (cm⁻¹): 3486, 3363, 1752, 1428, 1350, 1098, 1035; ¹H NMR (C₅D₅N, 400 MHz) and ¹³C NMR (C₅D₅N, 100 MHz) spectral data, see Table 1. FAB-MS (-) m/z: 371 [M - 1]⁻ (90). HR-ESI-MS (-): 371.1704 [M - 1]⁻ (calcd for C₁₈H₂₇O₈, 371.1705).

3.3.2 Crystallographic data for compound 1

 $C_{18}H_{28}O_8$, M = 372.40, orthorhombic, space group P2 (1), a = 6.5066 (14) Å, b = 34.978 (7) Å, c = 8.2676 (17) Å, $\beta = 90.00^{\circ}, V = 1881.6 (7) \text{ Å}^3, Z = 4.$ Crystal dimensions $(0.22 \times 0.20 \times$ 0.14 mm³) were used for the measurement on a MAC DIP-2030K diffractometer with a graphite monochromator $(\phi \text{ and } \omega \text{ scans}, 2\theta \max = 57.5^\circ),$ Mo K α radiation. The total number of independent reflections was 8407, of which 2882 were observed $(|F|^2 \ge 2)$ $\sigma |F|^2$). Final indices: $R_f = 0.0793$, $wR_2 = 0.1524$ ($w = 1/\sigma |F|^2$), S = 0.916. The crystal structure of compound 1 was solved by the direct method SHELXS-97 and expanded using geometrical calculations and difference Fourier techniques, refined by the program SHELXS-97 (Sheldrick, G. M. University of Gottingen: Gottingen, Germany, 1997) and the full-matrix least-squares calculations. Crystallographic data (excluding structure factors) for the structures in this paper have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication No. CCDC 656110. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (fax: +44 (0)1223 336033 or e-mail: deposit@ccdc.cam.ac.uk; http://www. ccdc.cam.ac.uk).

3.3.3 4-Pentylcyclohex-3-ene- 1α , 2β diol (2)

A colorless oil; $[\alpha]_D^{23} - 2.01$ (c = 0.74, MeOH); IR (KBr) ν_{max} (cm⁻¹): 3420, 1621, 1457, 1434, 1065, 1034, 909; ¹H NMR (C₅D₅N, 400 MHz) and ¹³C NMR (C₅D₅N, 100 MHz) spectral data, see Table 1. EI-MS *m*/*z*: 184 [M]⁺(5), 166 (2), 125 (13), 113 (13), 95 (18), 84 (100). HR-ESI-MS (+): 207.1353 $[M+Na]^+$ (calcd for $C_{11}H_{20}O_2Na$, 207.1360).

Acknowledgements

The authors are grateful to the staff of the analytical group of State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, for measurements of all spectra, and to Prof. Ming-Jin Xie (Yunnan University) for his professional measurement of the X-ray diffraction. This work was supported by the Natural Science Foundation of Yunnan (No. 312007C090M), the External Cooperation Program of Chinese Academy of Sciences (No. GJHZ200818), and CAS-Croucher Foundation (CAS-CF07/08.SC03).

References

- Chinese Pharmacopoeia Committee (Part 1), *Chinese Pharmacopoeia* (Chemical Industry Press, Beijing, 2005), p. 10.
- [2] Y.H. Li, S.L. Peng, Y. Zhou, K.B. Yu, and L.S. Ding, *Planta Med.* 72, 652 (2006).

- [3] L.S. Lim, P. Shen, Y.H. Gong, and E.L. Yong, *Phytochemistry* 67, 728 (2006).
- [4] Y.Q. Xiao, L. Li, X.L. You, M. Taniguchi, and K. Baba, *Chin. J. Chin. Mater. Med.* 27, 519 (2002).
- [5] T. Naito, Y. Ikeya, M. Okada, H. Mistuhashi, and M. Maruno, *Phytochem-istry* **41**, 233 (1996).
- [6] T. Naito, T. Katsuhara, K. Niitsu, Y. Ikeya, M. Okada, and L.H. Mitsuhashi, *Heterocycles* 32, 2433 (1991).
- [7] M. Kobayashi and H. Mitsuhashi, *Chem. Pharm. Bull.* 35, 4789 (1987).
- [8] M. Kaouadji, F. De Pachtere, C. Pouget, A.J. Chulia, and S. Lavaitte, *J. Nat. Prod.* 49, 872 (1986).
- [9] P. Wang, X. Gao, Y. Wang, Y. Fukuyama, I. Miura, and M. Sugawara, *Phytochem-istry* 23, 2033 (1984).
- [10] M. Kaouadji, H. Reutenauer, A.J. Chulia, and A. Marsura, *Tetrahedron Lett.* 24, 4677 (1983).
- [11] T. Ishikawa, T. Takayanagi, and J. Kitajima, *Chem. Pharm. Bull.* **50**, 1471 (2002).